# Ferrocene-based thiohydantoins: From synthesis to preliminarily *in vitro* antimicrobial/ anticancer activity - in search of new chemotherapeutic agents.

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### Abstract

In this work novel organic based compounds, nitrogen heterocycles were synthesized and their antimicrobial and anticancer activities. Anew series of nitrogen heterocycles (**3** and **7**) containing ferrocene ring were prepared via the reaction of 1-acetyl ferrocene (**1**) with thiosemicarbazide to give N-substituted thiourea (**2**), followed by cyclization with ethyl chloroacetate and phenacyl bromide. Condensation of **3** with aromatic aldehyde, yielded arylidene derivatives(**5**). Acetyation of compounds **3**, **5** and **7** with acetic anhydride gave the corresponding N-acetyl derivatives (**4**, **6** and **8**). The structures of the synthesized compounds were confirmed by IR, <sup>1</sup>H-NMR, <sup>13</sup>C-NMR, MS and elemental analysis. Antimicrobial and antitumor activities of some synthesized compounds have been investigated. The two compounds, selected as potential agents hepatocellular carcinoma (HCC) were then evaluated in vitro for their biological activity on HCC-derived cell lines(The compounds show a promising inhibitory growth efficacy (IC<sub>50</sub> = 2.61and 2-69µg) with compared standard antitumor drug (IC<sub>50</sub> = 4.60 µg).

Keywords: Nitrogen heterocycles, antimicrobial and anticancer activities.

# 1. Introduction

The hydantoin, imidazolidene-2, 4-dione, scaffold is an important structural component that is present in a number of natural products <sup>[1,2]</sup> and chemotherapeutic important compounds <sup>[3-5]</sup>. Hydantoins substituted at the C-5 position exhibit a wide range of pharmacological activities including antiarrhythmic and antihypertensive <sup>[6]</sup>, antiviral <sup>[7]</sup>, antineoplastic <sup>[8]</sup>, antitumoral <sup>[9]</sup>, anticonvulsant agents <sup>[10]</sup>, antidepressant <sup>[11]</sup> and platelet aggregation inhibitory activities <sup>[12]</sup>. Phenylhydantoins have attracted great interest because of their beneficial biological properties such as: antiepileptic <sup>[13]</sup>, herbicides <sup>[14]</sup> and fungicides <sup>[15]</sup>.

To the best of our knowledge, incorporation of ferrocene fragment into an organic compound often led to unexpected biological activity, which is due to their different membrane permeation properties and anomalous metabolism. Many ferrocenyl-based compounds display interesting cytotoxic, antitumor, antimalarial, antifungal and DNA-cleaving activity <sup>[16]</sup>. Recently, some new ferrocenyl-substituted heterocyclic compounds have been reported as potential pharmaceuticals <sup>[17, 18]</sup>. Moreover, the stability and non-toxicity of the ferrocenyl moiety is of particular interest rendering such drugs compatible with other treatment <sup>[19]</sup>.

In view of these observations and as a part of our ongoing program devoted to the synthesis of diverse heterocycles as antimicrobial/ anticancer agents <sup>[20, 21]</sup>, in this study, we report herein synthesis and preliminarily *in vitro* antimicrobial activity and cytotoxicity of thiohyantoin derivatives substituted with phenyl or arylidene and ferrocene moieties.

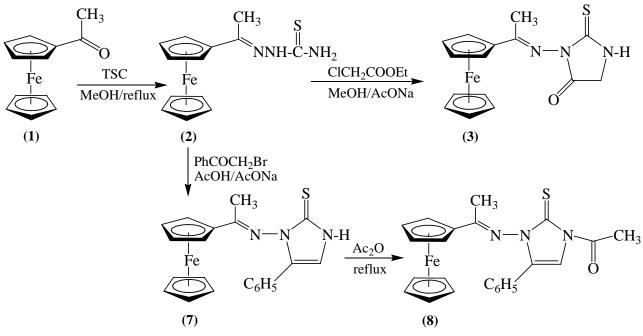
# 2. Results and discussion

# 2.1 Chemistry

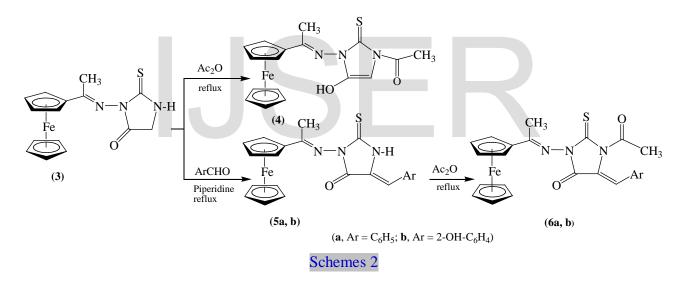
The synthesis of thiohyantoin-scaffold bearing phenyl or arylidene and ferrocenyl terminals (5-8) is depicted in Schemes 1, 2. The key starting materials 1-acetylferrocene thiosemicarbazone (2) was synthesized starting from 1-acetylferrocene (1) following modified literature procedure <sup>[3]</sup>, *via* refluxing of an ethanolic solution of 1-acetylferrocene and thiosemicarbazide. Treatment of the thiourea derivative (2) with ethyl chloroacetate in the presence catalytic amount of fused sodium acetate in methanol afforded the corresponding ferrocenyl-thiohyantoin, 3-[(ferrocene-1-ylethylidene)amino]-2-thioxo-imidazolidine-4-one (3) (Schemes 1). Furthermore, acetylferrocene thiosemicarbazone (2) undergoes coupling reaction with phenacyl bromide in the presence of fused sodium acetate in acetic acid under reflux to give the corresponding 4-phenyl-3-[(ferrocene-1-ylethylidene)amino]-imidazolidine-2-thione (7) which was converted to 1-acetyl-3-[(ferrocene-1-ylethylidene)amino]-4-phenyl-imidazolidine-2-thione (8) *via* the acylation with acetic anhydride under reflux conditions (cf. Schemes 1).

Ferrocenyl-thiohyantoin (**3**) was either converted into the 1-acetyl-3-[(ferrocene-1-ylethylidene) amino]-4-hydroxy-imidazolidine-2-thione (**4**) through boiling in acetic anhydride or reacted with two different aromatic aldehydes (**a**, **b**) in piperidine under fusion to yield the corresponding ferrocenyl-thiohyantoin arylidenes, 3-[(ferrocene-1-ylethylidene) amino]-2-thioxo-5-arylidene-imidazolidine-4-ones (**5a**, **b**) (see Scheme2). The ferrocenyl-thiohyantoin arylidenes (**5a**, **b**) were further acylated with acetic anhydride to afford the corresponding 1-acetyl-2-thioxo-3-[(ferrocene-1-ylethylidene)amino]-5-arylidene-imidazolidine-4-ones (**6a**, **b**).

International Journal of Scientific & Engineering Research, Volume 6, Issue 1, January-2015 ISSN 2229-5518



Schemes 1



# 2.2 Characterization of the Ferrocenyl-thiohyantoins

The ferrocenyl-thiohyantoins (**3-8**) were prepared in high yields, gave satisfactory C, H, N and S elemental analyses, which are consistent with the proposed formula. The IR spectra of compounds (**3-5**) exhibited characteristic peaks at 3210-3227, 1710-1728 and 1427-1462 cm<sup>-1</sup> due to thiohyantoin NH, carbonyl and thiocarbonyl moieties, respectively. Interestingly, the vibration at the region  $1632\pm5$  cm<sup>-1</sup> may be assigned to the imine (C=N) stretch. The NH stretches in the FTIR spectra of the - were missing from the spectra of compounds (**6**, **8**) indicating acylation of the thiohyantoin nitrogen and replacement of H by the acetyl group. This was further supported by the appearance of new strong peak at *ca*.  $1712\pm4$  cm<sup>-1</sup> in the spectra of (**6**, **8**), ascribed to acetyl C=O group.

# 2.3.1 Antibacterial activity;

Antibacterial activities were investigated using agar well diffusion method. The activity of tested samples were studied against the bacillus subtilis (RCMB 000 107) and staphylococcus aureus (RCMB 000 106) as gram positive bacteria, while Escherichia coli (RCMB 000 103) and pseudomonas aeruginosa (RCMB 000 102) as gram negative bacteria <sup>[22, 23]</sup>. The zone of inhibition was measured in mm. and was compared with standard drug. DMSO was used as a blank and ciprofloxacin was used as antibacterial standard. All the compounds were tested at 10 and 50 mg concentrations. The date is summarized in **table 1**, and show that all compounds display certain antibacterial activity.

		Gram post	itive bacteri	a	Gram negative bacteria				
Compd. No.	Bacillu	is Sub.	Staphyle	ococcus	E-C	Coli	Pseudomonas Sp.		
	10 mg	50 mg	0 mg 10 mg 50 mg 1		10 mg 50 mg		10 mg	50 mg	
2	-	4	6	10	-	7	2	9	
3	2	7	3	8	-	9	5	9	
4	-	1	-	2	-	4	-	1	
5a	9	13	8	14	8	14	9	15	
5b	5	12	6	11	7	10	4	11	
ба	6	10	7	13	7	13	5	12	
бb	5	11	5	10	4	10	6	11	
7	8	11	6	10	3	7	8	11	
8	7	9	3	8	5	9	7	10	
Ciprofloxacin	11	15	12	16	10	16	13	18	

Table 1: Antibacterial activity of nitrogen heterocycles

# 2.3.2 Antifungal activity;

Tested samples were screened separately using agar well diffusion method <sup>[22, 23]</sup> against various fungi (Aspergillus Nigaer and Penicillium Sp.). The inhibition zone was measured in mm. and was compared with fluconazole as standard drug, and using DMSO as a blank as antifungal activity. All the compounds were tested at 10 and 50 mg concentrations. The data are listed in **table 2**.

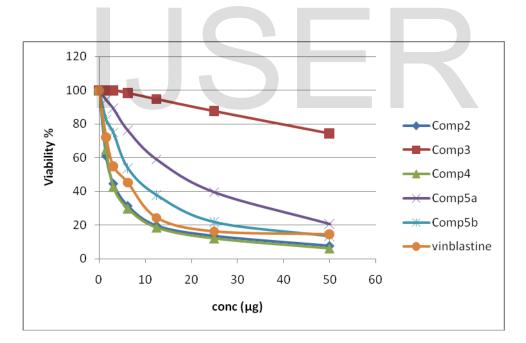
Compd. No.	Aspeargill	lus Nigaer	Penicllium Sp.			
Compa. 110.	10 mg	50 mg	10 mg	50 mg		
2	-	5	-	7		
3	2	8	-	9		
4	1	7	-	3		
5a	9	14	8	16		
5b	5	11	9	14		
6a	6	13	7	13		
6b	4	9	6	10		
7	5	10	6	11		
8	4	8	5	9		
Fluconazole	11	18	13	19		

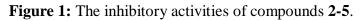
# 2.2.3. Cytotoxicity assay;

In this study, the anticancer activity of the **9** synthesized compounds bearing ferrocene moiety has been evaluated on human cancer cell lines, representing liver cancer. The cytotoxic activities of prepared compounds were tested against HepG-2 cell line according to method of Masmann and Vijayen et al <sup>[24, 25]</sup>. The inhibitory activity against liver carcinoma cells (HepG-2) was detected by using different concentrations of tested samples (50, 25, 12.5, 6.25, 3.125, and 1.65  $\mu$ g) and the viability cells (%) were determined by colorimetric method. The IC<sub>50</sub> was calculated from **table 3** and **figures 1** and **2**.

International Journal of Scientific & Engineering Research, Volume 6, Issue 1, January-2015 ISSN 2229-5518 **Table 3:** Evaluation of cytotoxicty of nitrogen heterocycles

Sample	Viability (%)									
Conc. (µg)	2	3	4	5a	5b	6a	6b	7	8	Vinblastine Standard
50	7.64	74.32	5.97	20.58	13.39	70.86	26.92	32.56	12.71	14.38
25	13.55	87.63	11.94	39.47	21.83	87.42	42.13	68.47	29.46	16.13
2.5	19.62	94.78	18.38	58.94	37.82	95.08	79.48	83.19	72.65	24.25
6.25	31.43	98.39	29.72	76.22	53.87	99.12	88.17	92.51	84.16	45.13
3.125	44.61	100.0	42.53	89.16	74.91	100.0	95.08	47.89	93.58	55.00
1.65	60.98	100.0	64.78	94.38	82.54	100.0	98.72	99.13	97.84	72.13
0.00	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.00





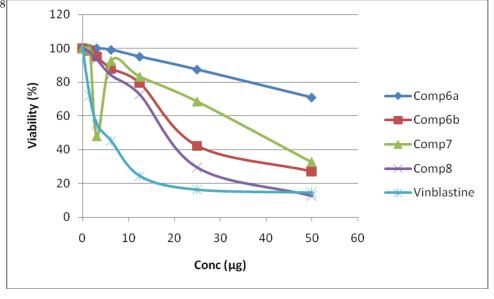


Figure 2: The inhibitory activities of compounds 6, 7 and 8.

The results of 50% inhibitory concentration (IC<sub>50</sub>) date are summarized in table 4.

Table 4: IC<sub>50</sub> (µg) values of nitrogen heterocycles after 24 h continuous exposure of tumor cell line

Compound No.	2	3	4	5a	5b	ба	6b	7	8	Vinblastine Standard
Tumor type/Cell line HepG-2	2.61	>50	2.69	18.20	7.76	>50	22.40	37.9	19.1	4.60

The  $IC_{50}$  value is the concentration that induces 50% growth inhibition compared with untreated control cells.

HepG-2: Human heputocellular carcinoma cell lines.

According to the results of cell culture studies, the compounds 2 and 4 showed antitumor activity than the standard antitumor drug against HepG-2 cell line.

In comparison with standard antitumor drug vinblastine, all another tested compounds have cytotoxic and antitumor activity less than standard antitumor drug against hepatocellular carcinoma cell line (HepG-2).

**3.** Conclusion

A new series of nitrogen heterocycles containing ferrocene ring were prepared. The structures of these compounds were confirmed by IR, <sup>1</sup>H-NMR, <sup>13</sup>C-NMR, MS and elemental analysis. Anticancer activities of synthesized compounds were evaluated on liver cancer cell lines. As a result of the cell culture studies, all of the compounds have shown anticancer activity for liver cancer cells. In conclusion, some nitrogen heterocycles compounds might be potentially useful in the field of cancer treatment, finally compounds **2**, **4** and **5b** can be suggested as potent candidates for liver cancer drug.

### 4. Experimental Section

## 4.1. Chemistry

All starting materials, reagents, and solvents were purchased from commercial suppliers. Melting points were measured in open capillary tubes on a MEL-TEMP II melting points apparatus and uncorrected. The NMR spectra were recorded on a BRUKER 300 BB instrument at room temperature, operating at 300 MHz for <sup>1</sup>H and 75.075 MHz for <sup>13</sup>C. The chemical shifts were recorded as  $\delta$  values in ppm units downfield of tetramethylsilane used as internal standard. The IR spectra were recorded on a FT/IR – 4200 spectrometer in KBr pellets. Mass spectra were obtained on a Joel JMS D-300 spectrometer operating at 70 eV. The elemental analyses were performed on a Perkin Elmer 2400 series II CHNS/O elemental analyzer.

# 1-[( Ferrocene-1-ylethylidene) amino]-thiourea (2)

A mixture of 2-acetylferrocine (1, 0.01mole) and thiosemicarbazide (0.01mole) in methanol (50 mL) was heated under reflux for 4 h, and then cooled. The solid formed was filtered off, dried and purified by recrystalization from methanol to give **2** as pale red crystals, yield 78%, m. p. 175°C. IR (KBr): 3410, 3128 (NH<sub>2</sub>), 3218 (NH), 1635 (C=N), 1431 (C=S) cm<sup>-1</sup>. <sup>1</sup>H-NMR (DMSO – d<sub>6</sub>):  $\delta$  2.19 (s, 3H, CH<sub>3</sub>), 3.32 (s, 2H, NH<sub>2</sub>), 4.17 – 4.80 (m, 9H, H–ferrocene ring), 9.95 (s, 1H, NH) ppm. <sup>13</sup>C-NMR (DMSO -d<sub>6</sub>):  $\delta$  178.42 (C = S), 151.03 (C = N), 83.55, 72.59, 71.16, 70.21, 70.10, 70.00, 69.83, 69.49, 68.19, 67.73 (C–ferrocine ring), 15.52 (CH<sub>3</sub>) ppm. MS: m/z (%) = 303 (M<sup>+</sup> + 2, 5.60), 302 (M<sup>+</sup> + 1, 18.30), 301 (M<sup>+</sup>, 79.00), 300 (M<sup>+</sup> - 1, 10.00), 285 (10.40), 284 (49.70), 283 (24.50), 269 (2.50), 268 (1.80), 267 (3.8), 266 (2.10), 259 (1.60), 258 (1.00), 249 (2.60), 248 (8.40), 247 (6.60), 237 (1.70), 236 (4.80), 235 (4.40), 234 (2.00), 228 (10.80), 227 (11.90), 226 (17.10), 219 (4.80), 218 (16.40), 217 (3.60), 212 (8.40), 211 (11.30), 202 (5.30), 201 (2.60), 197 (5.00), 196 (35.20), 195 (17.50), 194 (22.40), 193 (15.30), 186 (14.30), 185 (56.20), 184 (27.0), 178 (15.60), 177 (17.30), 176 (6.10), 161 (5.80), 160 (14.00), 159 (7.80), 154 (9.80), 153 (25.70), 152 (16.90), 151 (7.90), 148 (5.50), 147 (5.10), 146 (7.10), 138 (3.20), 137 (32.80), 136 (21.70), 135 (8.50), 129 (28.10), 128 (15.00), 127 (6.00), 122 (11.20), 121 (77.30), 120 (20.60), 119 (9.40), 107

(8.30), 106 (6.30), 105 (10.50), 104 (7.20), 97 (26.60), 96 (8.00), 95 (14.00), 94 (17.30), 89 (15.40), 88 (9.10), 81 (16.90), 80 (16.40), 74 (7.80), 77 (8.50), 66 (5.50), 65 (18.10), 64 (13.70), 63 (14.30), 57 (10.20), 56 (100), 55 (21.60), 54 (9.70), 51 (7.30). Anal. Found: C, 51.73; H, 4.89; N, 13.83; S, 10.48. C<sub>13</sub>H<sub>15</sub>N<sub>3</sub>FeS requires: C, 51.83; H, 4.98; N, 13.95; S, 10.63.

# 3-[(Ferrocene-1-ylethylidene) amino]-2-thioxo-imidazolidine-4-one (3)

A mixture of (2, 0.01mole), ethyl chloroacetate (0.01mole) and fused sodium acetate (0.03mole) in methanol (30 mL) was heated under reflux for 3-4 h. The solid formed after cooling was filtered off, washed with hot water, dried and purified by recrystallization from methanol to give **3** as pale red crystals, yield 73%, m. p. 240°C. IR (KBr): 3210 (NH), 1728 (C = O), 1627 (C = N), 1427 (C = S) cm<sup>-1</sup>. <sup>1</sup>H - NMR (DMSO – d<sub>6</sub>):  $\delta$  2.23 (s, 3H, CH<sub>3</sub>), 3.81 (s, 2H, NCH<sub>2</sub> CO), 4.17 – 4.68 (m, 9H, H-ferrocene ring), 10.30 (S, 1H, NH) ppm. <sup>13</sup>C–NMR (DMSO –  $d_6$ ):  $\delta$  178.11 (C = S), 169.31 (C = O), 151.31 (C = N), 83.24, 72.58, 70.41, 70.10, 69.78, 69.64, 67.70 (C - C)ferrocene ring),  $33.13(NCH_2CO)$ ,  $16.21(CH_3)$  ppm. MS: m/z(%)= 343 (M<sup>+</sup>+2, 8.70), 342 (M<sup>+</sup>+1, 23.10), 341 (M<sup>+</sup>, 100), 340 (M<sup>+</sup>-1, 22.40), 277 (4.50), 276 (16.40), 275 (8.70), 268 (3.10), 267 (5.20), 249 (5.90), 248 (11.50), 247 (10.80), 237 (5.90), 236 (15.00), 235 (5.60), 234 (15.00), 232 (5.90), 227 (7.30), 226 (8.70), 225 (4.50), 212 (7.00), 211 (8.70), 210 (5.60), 203 (7.70), 202 (34.30), 201 (18.20), 199 (7.30), 198 (3.80), 187 (5.90), 186 (10.10), 185 (23.10), 184 (11.50), 179 (5.2), 178 (5.60), 168 (5.60), 167 (3.50), 166 (11.50), 162 (22.40), 161 (14.00), 160 (9.80), 147 (8.40), 146 (9.40), 145 (6.60), 135 (5.90), 134 (11.50), 132 (7.70), 122 (4.50), 121 (53.50), 120 (9.80), 119 (8.40), 113 (11.90), 112 (4.90), 107 (5.90), 106 (5.90), 105 (7.70), 98 (2.80), 97 (11.20), 96 (6.30), 92 (5.60), 91 (14.00), 89 (5.20), 79 (3.50), 78 (8.00), 77 (8.70), 65 (13.90), 64 (12.90), 63 (8.70), 57 (2.80), 56 (56.60), 55 (9.10), 51 (5.60). Anal. Found: C, 52.67; H, 4.38; N, 12.28; S, 9.29.C<sub>15</sub>H<sub>15</sub>N<sub>3</sub>Fe OS requires: C, 52.78; H, 4.40; N, 12.32; S, 9.38.

# 5-Arylidene-3-[(ferrocene-1-ylethylidene)amino]-2-thioxo-imidazolidine-4-one (5a, b)

A mixture of (3, 0.01 mole), aromatic aldehydes (such as benzaldehyde and 4methoxybenzaldehyde) (0.01 mole) and piperidine (1mL) was fused on a hot-plate at 120-130°C for 1h. The reaction mixture was cooled and acidified with dilute hydrochloric acid (1N). The crude product was filtered off, washed with water, dried and purified by recrystallization from ethanol to give 5.

5-Benzylidene-3[(ferrocene-1-ylethylidene)amino]-2-thioxo-imidazolidine-4-one (5a) as brown crystals, yield 63%, m. p. 160°C. IR (KBr): 3221 (NH), 1710 (C = O), 1637 (C = N), 1605, 1589 (C = C), 1446 (C = S) cm<sup>-1</sup>. <sup>1</sup>H–NMR (DMSO– d<sub>6</sub>):  $\delta$  2.31 (s, 3H, CH<sub>3</sub>), 4.21 – 4.81 (m, 9H, H –

ferrocene ring), 7.44 – 7.88 (m, 6H, Ar- H and H – olefinic), 10.61 (s, 1H, NH) ppm.<sup>13</sup>C–NMR (DMSO – d<sub>6</sub>):  $\delta$  167.77 (C = S), 164.77 (C = O), 152.31 (C = N), 138.26, 136.64, 134.26, 130.29, 129.71, 128.86, 127.79, 124.06, 120.71 (C – aromatic and C – olefinic), 68.01, 69.75, 69.74, 69.89, 70.17, 70.26, 70.51, 70.75, 72.21, 82.82 (C – ferrocene ring), 16.56 (CH<sub>3</sub>) ppm. MS: m/z (%) = 430 (M<sup>+</sup> + 2, 11.30), 429(M<sup>+</sup> + 1, 9.30), 428 (M<sup>+</sup>, 17.35), 427(M<sup>+</sup>-1, 7.80), 404(6.30), 403(9.20), 227(8.30), 226(10.70), 225(3.50), 212(6.00), 211(7.80), 210(4.60), 203(8.70), 202(32.300, 201(17.20), 199(6.30), 198(4.80), 187(5.60), 186(9.10), 185(22.10), 184(10.50), 179(4.8), 178(6.60), 168(5.50), 167(3.30), 166(10.50), 162(20.40), 161(14.00), 160(7.80), 147(7.20), 146(8.30), 145(5.60), 135(17.20), 134(100), 132(6.80), 122(3.50), 121(50.60), 120(8.90), 119(8.30), 113(12.80), 112(3.80), 107(6.50), 106(4.90), 105(6.70), 98(2.30), 97(10.30), 96(5.30),92(5.60), 91(16.30), 90(34.20), 89(6.30), 79(3.30), 78(10.20), 77(42.50), 65(13.20), 64(17.10), 63(18.50), 57(3.60), 56(57.20), 55(8.10), 51(15.30). Anal. Found: C, 61.58; H, 4.01; N, 9.71; S, 7.37. C<sub>22</sub>H<sub>18</sub>N<sub>3</sub>FeOS requires: C, 61.68; H, 4.21; N, 9.81; S, 7.47.

5-(2-Hydroxy)benzylidene-3[(ferrocene-1-ylethylidene)amino]-2-thioxo-imidazolidine-4-one (5b) as brown crystals, yield 61%, m. p. 165°C. IR (KBr): 3227 (NH), 1713 (C = O), 1632 (C =N), 1608, 1593 (C = C), 1445 (C = S), 1095, 1052 (C = O) Cm<sup>-1</sup>. <sup>1</sup>H–NMR (DMSO – d<sub>6</sub>): δ 2.32 (s, 3H, CH<sub>3</sub>), 4.23 – 4.80 (m, 9H, H - ferrocene ring), 6.85 - 7.87 (m, 5H, Ar - H and H– olefinic), 10.71 (s, 1H, NH) and 11.35(s, 1H, OH) ppm. <sup>13</sup>C-NMR (DMSO – d<sub>6</sub>): δ 168.30 (C =S), 165.33 (C = O), 153.20 (C – O), 152.30 (N – C = N), 138.24, 136.63, 134.25, 130.28, 129.70, 128.83, 127.78, 120.73 (C – aromatic and C – olefinic), 82.80, 72.22, 70.52, 70.25, 70.16, 69.86, 69.73, 69.34, 68.02 (C – ferrocene ring), 16.53 (CH<sub>3</sub>) ppm. Anal. Found: C, 59.37; H, 3.95; N, 9.22; S, 7.03. C<sub>22</sub>H<sub>18</sub>N<sub>3</sub>FeO<sub>2</sub>S requires: C, 59.46; H, 4.05; N, 9.46; S, 7.21.

# 4-Phenyl-3[(ferrocene-1-ylethylidene)amino]-imidazolidine-2-thione (7)

A mixture of (**2**, 0.01mole), phenacyl bromide(0.01mole) and fused sodium acetate (0.03mole) in glacial acetic acid and (30 mL) was heated under reflux for 2-3 h, then cooled and poured into water. The resulting solid was filtered off, washed with water, dried and purified by recrystallization from ethanol to give **7** as red crystals, yield 64%, m. p.  $220^{\circ}$ C – IR (KBr): 3309 (NH), 1631 (C = N), 1602, 1585 (C = C), 1442 (C = S) cm<sup>-1</sup>, <sup>1</sup>H-NMR (DMSO – d<sub>6</sub>):  $\delta$  2.23 (s, 3H, CH<sub>3</sub>), 4.18 – 4.63 (m, 9H, H – ferrocene ring), 7.26 –7.88 (m, 6H, Ar – H and H – imidazolidine ring), 10.91 (s, 1H, NH) ppm.<sup>13</sup>C – NMR (DMSO - d<sub>6</sub>):  $\delta$  170.35 (C = S), 151.10 (N = C – N), 149.01 (=C– N), 135.44, 129.04, 127.84, 125.96, 103.98 (C–aramatic and C–imidazolidine), 84.43, 69.90, 69.45, 66.96 (C – ferrocene ring), 15.79 (CH<sub>3</sub>) ppm. MS: m/z (%) =403 (M<sup>+</sup> + 2, 7.50), 402 (M<sup>+</sup> + 1, 6.50), 401 (M<sup>+</sup>, 16.30), 387(6.20), 386(7.20), 358(6.10), 357(7.70), 228(11.80), 227(13.80), 226(19.300, 219(5.30), 218(17.30), 217(4.50), 212(6.40),

211(13.20), 202(5.10), 201(3.60), 197(5.30), 196(34.20), 195(18.20), 194(21.30), 193(16.20), 186(3.20), 185(56.20), 184(26.30), 178(15.60), 177(16.20), 175(7.20), 161(5.80), 160(13.00), 159(6.80), 154(6.80), 153(23.20), 152(16.80), 151(6.90), 148(4.90), 147(5.20), 146(8.20), 138(2.20), 137(31.20), 136(20.80), 132(10.10), 131(18.50), 128(13.10), 127(5.50), 122(10.50), 121(76.20), 120(14.60), 119(8.30), 107(6.30), 106(5.80), 105(11.30), 104(7.30), 103(23.20), 102(11.70), 92(11.30), 91(17.20), 89(61.30), 88(6.10), 78(7.80), 77(18.50), 76(11.20), 66(4.50),65(11.30), 64(11.20), 63(22.20), 57(11.30), 56(100), 55(23.60), 54(8,70), 51(7.20). Anal. Found: C, 62.73; H, 4.64; N, 10.33; S, 7.83. C<sub>21</sub>H<sub>19</sub>N<sub>3</sub>FeS requires: C, 62.84; H, 4.74; N, 10.47; S, 7.98.

1-Acetyl-3-[(ferrocene-1-ylethylidene) amino]-4-hydroxy imidazolidine-2-thione (4)
1-Acetyl-2-thioxo-3-[(ferrocene-1-ylethylidene) amino]-5-arylidine- imidazolidine-4-ones (6a, b)
1-Acetyl-3-[(ferrocene-1-ylethylidene) amino]-4-phenyl-imidazolidine-2-thione (8)

A solution of **3**, **5** and **7** (0.01mole) in acetic anhydride (20 ml) was heated under reflux for 2 hr, then cooled and poured into ice-water. The resulting solid was filtered off, washed with water, dried and purified by recrystallization from benzene to give **4**, **6** and **8**.

*Compound 4* as pale red crystals, yield 58%, m. p. 150°C. IR (KBr): 3460 - 2980 (br. OH), 1728 (C = O), 1635 (C = N), 1462 (C = S), 1221, 1095 (C = O) cm<sup>-1</sup>. <sup>1</sup>H-NMR (DMSO - d<sub>6</sub>):  $\delta$  2.13 (s, 3H, COCH<sub>3</sub>), 2.24 (s, 3H, N = C - CH<sub>3</sub>), 3.32 (br. s, OH), 4.14 - 4.64 (m, 10H, H - ferrocene ring and H - imidazolidine) ppm.

<sup>13</sup>C-NMR (DMSO – d<sub>6</sub>):  $\delta$  165.85 (C = S), 164.24 (C = O), 151.30 (-C = N), 150.23 (C – OH), 91.45 (CH of imidazolidine), 63.96, 68.07, 67.92, 67.27, 67.26 (C – ferrocene ring), 23.40, 23.16 (2 × CH<sub>3</sub>) ppm. MS:m/z (%) = 385 (M<sup>+</sup> + 2, 7.80), 384 (M<sup>+</sup> + 1, 21.30), 383 (M<sup>+</sup>, 32.30), 343 (6.80), 342 (36.20), 341 (100), 340 (21.40), 277 (4.30), 276 (16.50), 257 (8.30), 268 (3.11), 267 (5.01), 249 (5.80), 247 (0.78), 237 (5.70), 236 (15.30), 234 (15.01), 232 (5.80), 227 (7.31), 226 (8.80), 225 (4.60), 212 (8.10), 211 (8.80), 210 (6.50), 203 (7.60), 202 (34.20), 201 (18.10), 199 (7.20), 198 (3.83), 187 (5.80), 186 (10.98), 185 (23.00), 184 (11.40), 179 (5.50), 178 (5.65), 168 (5.50), 167 (3.50), 166 (11.30), 162 (22.30), 161 (13.80), 160 (9.83), 147 (8.33), 146 (9.40), 145 (6.60), 135 (5.85), 134 (11.40), 132 (7.68), 122 (4.50), 121 (53.30), 120 (9.60), 118 (8.30), 113 (11.80), 112 (4.40), 107 (5.80), 106 (5.80), 105 (7.76), 98 (2.70), 97 (11.30), 96 (6.30), 92 (5.50), 91 (14.30), 89 (5.30), 79 (3.40), 78 (8.10), 77 (8.70), 65 (13.80), 64 (12.90), 63 (8.80), 57 (2.80), 56 (56.30), 55 (9.20), 51 (5.63). Anal. Found: C, 53.16; H, 4.22; N, 10.78; S, 8.27. C<sub>17</sub>H<sub>17</sub>N<sub>3</sub>FeO<sub>2</sub>S requires: C, 53.26; H, 4.44; N, 10.96; S, 8.35.

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Compound 6a as pale brown crystals, yield 63%, m. p.  $120^{\circ}$ C. IR (KBr): 1720 - 1712 (br. C = O), 1628 (C = N), 1468 (C = S), 1605, 1585 (C = C) cm<sup>-1</sup>.<sup>1</sup>H–NMR (DMSO – d<sub>6</sub>):  $\delta$  2.18 (s, 3H, CO CH<sub>3</sub>), 2.34 (s, 3H, N = C-CH<sub>3</sub>), 4.16 – 4.47 (m, 9H, H – ferrocene ring), 6.68 – 7.81 (m, 6H, Ar-H and H-olefinic) ppm.  ${}^{13}C$  – NMR (DMSO – d<sub>6</sub>):  $\delta$  166.39, 158.33 (C = O), 167.20 (C = S), 146.31 (C = N), 133.24, 132.17, 130.89, 130.05, 129.53, 127.69, 125.19, 91.18 (C-aromatic and Colefinic), 84.84, 64.40, 69.80, 69.54, 69.47, 68.84, 68.38, 67.95, 67.42, 67.36 (C-ferrocene ring), 23.53, 23.30 (2 × CH<sub>3</sub>) ppm. MS: m/z (%) = 472(M<sup>+</sup> +2, 6.30), 471(3.50), 470(M<sup>+</sup>, 9.20), 430(6.20), 429(7.70), 428(100), 427(13.20), 40495.30), 403(11.60), 228(1.30), 227(11.50), 226(11.80), 225(4.30), 212(7.20), 211(8.30), 210(5.50), 203(7.80), 202(31.20), 201(15.80), 199(5.40), 198(6.70), 187(3.60), 186(8.10), 185(32.10), 184(11.30), 179(4.50), 178(6.30), 168(6.20), 167(4.20), 166(11.30), 162(19.20), 161(11.30), 160(6.70), 147(5.70), 146(6.60), 145(6.50), 135(17.10), 134(86.50), 133(11.20), 132(6.80), 122(4.30), 121(35.60), 120(6.90), 119(7.30), 113(11.20), 112(4.20), 107(6.30), 106(5.20), 105(7.20), 98(2.10), 97(11.20), 92(6.50), 105(7.20),91(17.20), 90(33.20), 89(16.20), 78(3.30), 77(45.20), 65(11.20), 64(9.80), 63(22.30), 57(11.20), 56(69.30), 55(8.20), 51(11.20). Anal. Found: C, 61.11; H, 4.09; N, 8.78; S, 6.66. C<sub>24</sub>H<sub>20</sub>N<sub>3</sub>FeO<sub>2</sub>S requires: C, 61.27; H, 4.25; N, 8.43; S, 6.81.

*Compound 6b* as pale brown crystals, yield 61%, m. p. 137°C. IR (KBr): 1722 – 1715 (br. C = O), 1632 (C = N), 1605, 1589 (C = C), 1463 (C = S), 1215, 1078 (C – O) cm<sup>-1</sup>.<sup>1</sup>H–NHR (DMSO–d<sub>6</sub>):  $\delta$  2.21 (s, 3H, COCH<sub>3</sub>), 2.33 (s, 3H, N = C–CH<sub>3</sub>), 4.15 – 4.73 (m, 9H, H – ferrocene ring), 6.83 – 7.81 (m, 5H, Ar–H and H–olefinic) ppm. <sup>13</sup>C–NMR (DMSO – d<sub>6</sub>):  $\delta$  168.20, 166.30 (C = O), 165.31 (C =S), 152.20 (C – O), 151.12 (C = N), 138.23, 136.62, 134.15, 130.26, 129.68, 128.71, 127.68, 120.61 (C –aromatic and C –olefinic), 83.80, 72.22, 70.51, 70.21, 70.15, 69.83, 69.72, 69.33, 68.11 (C – ferrocene ring), 23.18, 23.51 (2×CH<sub>3</sub>), 21.21(CH<sub>3</sub>) ppm. Anal. Found: C, 58.98; 4.02; N, 7.79; S, 6.02. C<sub>26</sub>H<sub>22</sub>N<sub>3</sub>FeO<sub>3</sub>S requires: C, 59.09; H, 4.17; N, 7.95; S, 6.06.

*Compound* 8 as pale brown crystals, yield 76%, m. p. 79 °C IR (KBr): 1708 (C = O), 1630 (C = N), 1605, 1589 (C = C), 1462 (C =S) cm<sup>-1</sup>.<sup>1</sup>H–NMR (DMSO – d<sub>6</sub>):  $\delta$  2.13 (s, 3H, COCH<sub>3</sub>), 2.37 (s, 3H, N = C–CH<sub>3</sub>), 4.12 – 4.79 (m, 9H, H – ferrocene), 7.25 – 7.88 (m, 6H, Ar–H and H– imidazolidine) ppm. <sup>13</sup>C – NMR (DMSO – d<sub>6</sub>):  $\delta$  180.39 (C = S), 168.89 (C = O), 149.22 (C = N), 134.74, 129.28, 129.06, 128.53, 128.24, 126.33, 126.13, 109.97 (C–aromatic and C – imidazolidine), 80.04, 71.74, 70.38, 70.02, 69.24, 68.97, 68.58 (C – ferrocene), 22.87 (N = C-CH<sub>3</sub>), 18.06 (CO -CH<sub>3</sub>) ppm. MS: m/z (%) = 445(M<sup>+</sup> +2, 6.70), 444(M<sup>+</sup>+1, 3.20), 443(M<sup>+</sup>,16.20), 403(6.50), 402(7.20), 401(100), 387(5.50), 386(9.50), 357(5.60), 228(11.60), 227(12.50), 226(21.20), 219(6.30), 218(18.30), 217(6.30), 212(5.40), 211(14.20), 202(4.70), 201(3.30), 197(6.10), 196(29.30), 195(16.50), 194(19.70), 193(17.20), 186(2.30), 185(40.10), 184(17.20), 178(11.20), 177(15.30), 175(6.30), 161(6.20), 160(13.20), 159(6.20), 154(7.20), 153(22.30), 152(15.60), 151(5.90), 148(3.90), 147(4.20), 146(7.30), 138(3.10), 137(30.30), 136(19.70), 132(9.20), 131(7.50), 128(11.30), 127(6.30), 122(11.60), 121(52.20), 120(18.20), 119(6.70), 107(5.60), 106(4.90), 105(9.70), 104(6.20), 103(21.30), 102(12.30), 92(12.60), 91(21.20), 89(15.60), 89(17.60), 88(7.10), 78(9.90), 77(21.20), 76(17.30), 66(5.40), 65(17.20), 64(16.50), 63(32.30), 57(17.20), 56(67.20), 55(11.20), 54(11.20), 51(18.30). Anal. Found: C, 62.33; H, 4.57; N, 9.29; S, 7.07.  $C_{23}H_{21}N_{3}FeO_{4}S$  requires: C, 62.30; H, 4.74; N, 4.48; S, 7.22.

# 4.2. Biological activity

# 4.2.1. Antibacterial assay of nitrogen heterocycles

Centrifuged pellets of bacteria from a 24 h. old culture containing approximatelly 104 - 106 CFU (colony forming unit) per ml. were sepread on the surface of nutrient agar (typtone 1%, yeast extrcat 0.5%, NaCl 0.5%, agar 1%, 1000 ml. of distilled water, PH 7.00), which was autoclaved under 121°C for at least 20 min. Wells were created in medium with the help of a strile metallic bores and then cooled down to 45°C. The activity was determined by measuring the diameter of the inhibition zone (in mm.).100  $\mu$ l of the tested samples (10 mg/ml and 50 mg/ml) were loaded into the wells of the plates. All compounds were prepared in dimethylsulfoxid (DMSO), DMSO was loaded as control. The plates were kept for incubation at 37°C for 24 h and then the plates were examined for the formation of zone of inhibition. Each inhibition zone was measured three times by caliper to get an average value. The test was preformed three times for each bacteria culture. Ciprofloxacin was used as antibacterial standard drug.

# 4.2.2. Antifungal evaluation of nitrogen heterocycles

Fungal strain was grown in 5 ml. sabourad dextrose broth (glucose: peptone; 40.10) for 3-4 days to achieve 105 CFU/ml cells. The fungal (culture co. 1 ml) was sepread out uniformly in the sabourad dextrose agar plates by sterilized triangular folded glass rod. Plates were left for 5-10 min. So that culture is properly adsorbed on that surface of sabourad dextrose agar plates. Now small wells of size (4 mm  $\times$  2 mm) were cut into the plates with the help of well culture and bottom of the wells were seald with 0.8% soft agar to prevent the flow of test sample at the bottom of the well.100  $\mu$ l of the tested samples (10 mg/ml and 50 mg/ml) were loaded into the wells of the plates .All compounds were prepared in dimethylsulfoxide (DMSO), DMSO was loaded as control. The plates were kept for incubation at 30°C for 3-4 days and then the plates were examined for the formation of zone of inhibition. Each inhibition zone was measured three times by caliper to get an average value. The test was performed three times for each fungus. Flucomazole was used as antifungal standard drug.

### 4.2.3. Cytotoxic assay of nitrogen heterocycles

# **4.2.3.1.** *Cell lines and culture*

Human hepatocellular carcinoma (HepG-2) cells were obtained from the American Type Culture Collection (ATCC, Rock Villa, MD). The cells were grown on RPMI – 1640 medium supplemented with 10% inactivated fetal calf serum and 50  $\mu$ g/ml gentamycin. The cells were maintained at 37°C in a humidified atmosphere with 5% CO<sub>2</sub> and were subcultured two to three times a week.

### **4.2.3.2.** Evaluation of the antitumor activity

The antitumor activity was evaluated on HepG-2 cell. The cells were grown as monolayer in growth RPMI – 1640 medium supplemented with 10% inactivated fetal calf serum and 50 µg/ml gentamycin. The monolayer of 10.000 cells adhered at the bottom of the wells in a 96-well microtiter plate incubated for 24 h at 37°C in a humidified incubator with 5% CO<sub>2</sub>. The monolayers were then washed with sterile phosphate buffered saline (0.01M, PH 7.20) and simultaneously the cells were treated with 100 µl from different dilution of the test sample in fresh maintenance medium and incubated at 37°C. A control of untreated cells was made in the absence of the test sample. Six wells were used for each concentration of the test sample. Every 24 h the observation under the inverted microscope was made. The number of the surviving cells was determined by staining the cells with crystal violet followed by cell lysing using 33% glacial acetic acid and read the absorbance at 490 nm using ELISA reader (SunRise, TECAN, Inc, USA) after well mixing. The absorbance values from untreated cells were considered as 100% proliferation. The number of viable cells was determined using ELISA reader as previously mentioned before and the percentage of viability was calculated as  $[1 - (ODt /OD_C)]$  100%, were ODt is the mean optical density of wells treated with the test sample and  $OD_C$  is the mean optical density of untreated cells. The 50% inhibitor concentration (IC<sub>50</sub>), the concentration required to cause toxic effects in 50% of intact cells, was estimated from graphic plots.

# Aknowledgements

We deeply appreciate the regional center for mycology and biotechnology Al-Zahar University for helping on the evaluation of cytotoxicit-y against liver cancer cells and antimicrobial activity.

### References

- [1] (a) M. Meusel, M. Gutschow, Org. Prep. Proced. Int., 2004, 36, 391; (b) L. Chevolot, S. Padua,
- B. N. Ravi, P. C. Blyth and P. J. Scheuer, *Heterocycles*, **1977**, **7**, 891; (c) R. Fathiafshar and T. M. Allen, *Can. J. Chem.*, **1988**, **66**, 45.
- [2] (a) C. Jimenez, P. Crews, *Tetrahedron Lett.*, 1994, 35, 1375; (b) I. S. Chen, C. T. Chang, W. S. Sheen, C. M. Teng, I. L. Tsai, C. Y. Duh, F. N. Ko, Phytochemistry, 1996, 41, 525.
- [3] C.-J. Fang, G. Han, Y.-J. Liu, C.-Y. Duan and Q.-J. Meng, Acta Cryst. (1999). C55, 2058-2060
- [4] (a) A. Balog, M.E. Salvati, W.F. Shan, A. Mathur, L.W. Leith, D.D. Wei, R.M. Attar, J.P. Geng, C. A. Rizzo, C. H. Wang, S. R. Krystek, J. S. Tokarski, J.T. Hunt, M. Gottardis, R. Weinmann, *Bioorg. Med. Chem. Lett.*, 2004, 14, 6107 (b) X.Q. Zhang, G.F. Allan, T. Sbriscia, O. Linton, S.G. Lundeen and Z. H. Sui, *Bioorg. Med. Chem. Lett.*, 2006, 16, 5763; (c) G.G. Muccioli, D. Martin, G.K.E. Scriba, W. Poppitz, J.H. Poupaert, J. Wouters, D.M. Lambert, *J. Med. Chem.*, 2005, 48, 2509.
- [5] T. Dylag, M. Zygmunt, D. maciag, J. handzlik, M. Bednarski, B. Filipek, K. Kiec-Kononowiez, *Eur. J. Med. Chem.*, **39**, 1013 – 1027, (**2004**).
- [6] N. Opacic, M. Barbaric, B. Zorc, M. Cetina, A. Nagl, D. Erkouic, M. Kradj, K. Pavelic, J. Balzarini, G. Andrei, R. Snoeck, E. De Clerey, S. Raic-Malic, M. Mintas, *Med. Chem.*, 48, 475-482, (2005).
- [7] E. Latt Mann, W. O. Ayuko, D. Kinchinaton, C. A. Langley, H. Singh, L. Karimi, M. S. Trsdale, *J.Phar.pharmacol*, 55, 1259-1265, (2003).
- [8] C. Carmi, A. Cavazzoni, V. Zuliani, A. Lodola, F. Bordi, P.V. Plazzi, R.R. Alfieri, P.G. Petronini, M. Mor, *Bioorg. Med. Chem. Lett.*, 16, 4021-4025, (2006).
- [9] G. Singh, P.H. Driver, J.W. Sander, L. Sander, *Brain*, 128, 7-17, (2005).
- [10] F. L. Wessels, T. J. Schwan and S. F. Pong, J. Pharm. Sci., 1980, 69, 1102.
- [11] A.G. Caldwell, C.J. Harris, R. Stepney and N. Whittaker, J. Chem. Soc., Perkin Trans. 1, 1980,495.
- [12] H.H. Merrit, T.J. Putnam, Arech. Neurol. Psychiatry, 39, 1003-1015, (1938).
- [13] M. Shiozaki, carbohydr. Res., 337, 2077-2088,(2002).
- [14] J. Marton, J. Fnisz, S. Hosztafi, T. Timar, J. Agric. Food. Chem, 41, 148-152, (1993).

- [15] (a) T.J. Kealy, P.L. Pauson, Nature 168 (1951) 1039-1040; (b) P.N. Kelly, A. Pretre, S. Devoy, I. O'Rielly, R. Devery, A. Goel, J.F. Gallagher, A. J. Lough, P.T.M. Kenny, *J. Organomet. Chem.* 692 (2007) 1327-1331; (c) M.F.R. Fouda, M.M. Abd-Elzaher, R.A. Abdelsamaia, A.A. Labib, *Appl. Organomet. Chem.* 21 (2007) 613-625.
- [16] (a) H. Yu, L. Shao, J. Fang, J. Organomet. Chem. 692 (2007) 991-996; (b) M. Zora, M. Görmen, J. Organomet. Chem. 692 (2007) 5026-5032. (c) M. Zora, O. Velioglu, J. Organomet. Chem. 693 (2008) 2159-2162.
- [17] (a) B. Fabian, V. Kudar, A. Csampai, T. Zs. Nagy, P. Sohar, J. Organomet. Chem. 692 (2007) 5621-5632; (b) T. Mochida, F. Shimizu, H. Shimizu, K. Okazawa, F. Sato, D. Kuwahara, J. Organomet. Chem. 692(2007)1834e1844. [28] B. Maity, M. Roy, A. R. Chakravarty, J. Organomet. Chem. 693 (2008) 1395-1399.
- [18] (a) C. Biot, L. Delhaes, L.A. Maciejewaski, M. Mortuaire, D. Camus, D. Dive, J.S. Brocard, *Eur. J. Med. Chem.* 35 (2000) 707-714; (b) C. Biot, G.Glorin, L.A. Maciejewaski, J. S. Brocard, *J. Med. Chem.* 40 (1997) 3715-3718; (c) C. Biot, L. Delhaes, L.A. Maciejewaski, D. Camus, D. Dive, J.S. Brocard, *Bioorg. Med. Chem.* 7 (1999) 2843-2847.
- [19] (a) I.M. El Deen, J.A. Hasanen, M. El Ashery, Inter. J. Innov. Res. In Science, Eng. And Technology, 3, 9702-9722, (2014); (b) H.K. Ibrahim, J.A. Hasanen, M.A. Zein and I.M. El Deen, Mens Agitat, 3, 54-72, (2008); (c) J.A. Hasanen, I.M. El-Deen, R.M. El-Desoky, A.M Abdalla, Res. Chem. Intermed. 2014, 40 (2) 537-553; (d) M.S. Refat, I.M. El-Deen, R.F.M. El-Shaarawy, Russ. J. Gen. Chem. 2014, 84 (3) 593-601.
- [20] (a) H. K.Ibrahim, E. Eltamany, R.F.M.Elshaarawy, I.M. Eldeen, *Maced. J. Chem. Chem. Eng.* 2008, 27(1), 65-79; (b) R.F.M. Elshaarawy, C. Janiak, *Eur. J. Med. Chem.* 2014, 75, 31-42; (c) R.F.M. Elshaarawy, Z.H. Kheiralla, A.A. Rushdy, C. Janiak, *Inorg. Chim. Acta* 2014, 421,110-122; (d) .F.M. Elshaarawy, C. Janiak, *Tetrahedron*, 2014, in press.
- [21] C.-J. Fang, G. Han, Y.-J. Liu, C.-Y. Duan and Q.-J. Meng, Acta Cryst. (1999). C55, 2058-2060.

[22] K.E. Cooper, A. Kavanagh, "Analytical microbiology", Vol-2 (Academic Press New York), 13, (1972).

[23] V. Betina, The chemistry and biology of antibiotres (Ed<sup>1</sup>) with nature RF Rekker, Cechoslovakia (*Elsevier Scientific Publishing, New York*), (**1983**).

[24] T. Mosmann, "Rapid Colorimetric Assay for Cellular Growth and Survival: Application to Proliferation and Cytotoricty Assays", *J. Immunol. Methods*, **65**, 55-63, (**1983**).

[24] N. Gangadevi, J. Muthumary, "Preliminary Studies on Cytotoxic effect of Fungal taxol on Cancer cell lines", *African J/ Biotechnology*, **6**, 1382-1386, (**2007**).

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